

Cell Invasion Assay Kit

(BME, 8 μ m, 24-well)

Instruction Manual

Catalog Number	PK-CA577-K913																							
Description	<p>Cell invasion is the ability of cells to migrate from one area to another through an extracellular matrix. Cell invasion is exhibited by both normal cells as well as cancerous cells in response to specific external signals, including chemical & mechanical stimuli. During invasion, extracellular matrix is enzymatically degraded by cellular proteases before cells migrate to the new location. Cell invasion is required for normal processes such as wound repair, vasculature formation and the inflammatory response as well as the abnormal invasion of tissues by tumor cells during metastasis.</p> <p>PromoCell's <i>Cell Invasion Assay Kit</i> (BME, 8 μm, 24-well) utilizes a Boyden chamber coated with Basement Membrane Extract (BME), where the cells invade the matrix and then migrate through a semipermeable membrane in the Boyden chamber in response to stimulants or inhibitory compounds. The percent cell invasion can be analyzed directly in a plate reader. Our assay is easy to use, sensitive and adaptable to high-throughput systems.</p>																							
Quantity	12 assays																							
Applications	<ul style="list-style-type: none"> • Measure cell invasion in response to stimuli • Screen and characterize compounds that influence cell invasion 																							
Sample Type	<ul style="list-style-type: none"> • Invasive cell lines • Invasion inhibitor or stimuli 																							
Kit Components	<table border="1"> <thead> <tr> <th>Components</th> <th>Quantity</th> <th>Cap code</th> </tr> </thead> <tbody> <tr> <td>Cell Invasion Chamber</td> <td>1 each</td> <td>Plate</td> </tr> <tr> <td>Wash Buffer</td> <td>25 ml</td> <td>NM</td> </tr> <tr> <td>Basement Membrane Solution</td> <td>1.2 ml</td> <td>Green</td> </tr> <tr> <td>Control Invasion Inducer</td> <td>300 μl</td> <td>Red</td> </tr> <tr> <td>Cell Dissociation Solution</td> <td>6 ml</td> <td>NM</td> </tr> <tr> <td>Cell Dye</td> <td>1.5 ml</td> <td>Blue</td> </tr> </tbody> </table>	Components	Quantity	Cap code	Cell Invasion Chamber	1 each	Plate	Wash Buffer	25 ml	NM	Basement Membrane Solution	1.2 ml	Green	Control Invasion Inducer	300 μ l	Red	Cell Dissociation Solution	6 ml	NM	Cell Dye	1.5 ml	Blue		
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User Supplied Reagents & Equipment	<ul style="list-style-type: none"> • Fluorescence Plate Reader • Cell Culture Media • Cotton Swabs • Centrifuge to spin 96-well plate • 96-well clear bottom white plate <p>For human primary cells and the corresponding optimized cell culture media as well as Cytokines/Chemokines and Growth Factors visit www.promocell.com.</p>																							
Applications / Assay Protocol	<p>A. General Consideration and Reagent Preparation:</p> <p>Store kit at -20°C, protected from light. Briefly centrifuge small vials prior to opening. Assay is performed under sterile conditions. Read entire protocol before performing the experiment.</p> <p>Cell Invasion Chamber: Open under sterile conditions. Keep at room temperature.</p> <p>Basement Membrane Solution: Thaw vial prior to experiment slowly on ice or in frost-free 4°C refrigerator. Temperatures above 4°C will rapidly turn the Base Membrane Solution into a gel. Thawing may take overnight at 4°C. The thawed matrix can be stored at 2-8°C for one week.</p> <p>For long term (6 months) storage, we recommend aliquoting into several tubes according to use and storing at -20°C.</p> <p>Control Invasion Inducer, Cell Dissociation Solution & Wash Buffer: Store at -20°C. Bring to 37°C before use. Stable for six months.</p> <p>Cell Dye: Aliquot and store at -20°C. Bring to 37°C before use.</p> <p>B. Cell Invasion Assay Protocol:</p> <ol style="list-style-type: none"> 1. Add 100 μl of Basement Membrane Solution to coat desired wells of the Top Chamber. Incubate plate at 37°C in an incubator for 1 hour to gel the Basement Membrane Solution. 2. Grow cells of interest in desired media and culture conditions. Grow enough cells to perform a 																							

Cell Invasion Assay and a Standard Curve. Adherent cells should be cultured to ~80% confluence.

3. Prior to the assay, starve cells for 18-24 hours in a serum-free media (0.5% serum can be used, if needed). After starvation, harvest the cells and centrifuge at 1,000 x g, for 5 minutes to pellet cells. Resuspend cell pellet in Wash Buffer and count the number of cells using a hemocytometer or an automated cell counter. Resuspend cells at 1×10^6 cells/ml in a serum-free media.

4. Under sterile conditions, disassemble the Cell Invasion Chamber and carefully remove the plate cover and the top chamber.

Bottom Chamber: Add 600 μ l of medium per well containing the desired chemoattractant to the bottom chamber. In control well(s), we recommend omitting the chemoattractant. For Positive Control, add 60 μ l of Control Invasion Inducer to 540 μ l of medium in the bottom chamber. Reassemble the top and bottom chambers while ensuring no air bubbles are trapped between them.

Top Chamber: Add 200 μ l ($2-3 \times 10^5$ cells) of cell suspension to each well of the top chamber. Add desired stimulator or inhibitor to the top well, and gently mix. Carefully replace the plate cover and incubate the Cell Invasion Chamber at 37°C in CO₂ incubator for 2-48 hours.

Note: Invasive cells pass through the basement membrane and cling to the outer side of the top chamber. Non-invasive cells stay in the upper chamber.

5. Standard Curve:

A. Each cell type requires a separate Standard Curve. Prepare a Standard Curve by adding 50 μ l cells (1×10^6 cells/ml, ~50,000 cells) in desired well(s) in a 96-well plate (white plate clear bottom). Serially dilute the cells 1:1 in Wash Buffer and generate a Standard Curve with different cell numbers (50,000; 25,000; 12,500; 6,250; 3,125; 1,562; 781 and 390) in 100 μ l total volume. As blank, use 100 μ l of Wash Buffer.

B. Add 10 μ l of Cell Dye to each well. Incubate at 37°C for 1 hour. Read the fluorescence at Em/Ex 530/590 nm. Plot the Standard Curve (Number of Cells vs RFU obtained). Fit the data points using a linear trend line with zero intercept. The equation for the straight line and R-squared value are used for data analysis of samples.

Note: The Cell Invasion RFU reading should fall in the linear range of the Standard Curve. We recommend using triplicates for Standard Curve.

6. Separation of Invasive Cells:

A. After the desired incubation with cell invasion inducers/inhibitors, carefully remove the plate cover and aspirate media from the top chamber without puncturing the membrane and matrix.

B. Remove cells from the top chamber using a cotton swab. Disassemble the Cell Invasion Chamber by removing the top chamber. Invert the top chamber and set it aside.

C. Place the plate cover on top of bottom chamber and centrifuge the plate at 1,000 x g for 5 minutes at room temperature.

D. Carefully aspirate the media from the bottom chamber and wash the chamber with 500 μ l Wash Buffer.

E. Centrifuge the plate at 1,000 x g for 5 minutes at room temperature and aspirate the Wash Buffer from the bottom chamber.

7. Count Invasive Cells:

A. For every twenty wells to be assayed, prepare a mix of 1 ml of Cell Dye in 10 ml of Cell Dissociation Solution. Mix well. Make the Cell Dye solution as desired depending on the number of wells.

B. Add 550 μ l of the mix to each well of the bottom chamber. Reassemble the Cell Invasion Chamber by placing the top chamber into the bottom chamber. Incubate at 37°C in a CO₂ incubator for 60 minutes*.

C. After incubation, disassemble the Cell Invasion Chamber, remove the top chamber and transfer 110 μ l of mix from the bottom chamber to the 96-well white plate (the same plate having Standards).

D. Read the plate at Ex/Em = 530/590 nm. Multiply the reading by 5 to account for the 5X higher volume in each well of the 24-well plate. Calculate the number of cells invaded using the equation of the straight line obtained from Standard Curve. Percentage Invasion can be calculated as follows:

Calculation:

$$\% \text{ Invasion} = \frac{\text{Number of Cells in Lower Chamber}}{\text{Total Number of Cells added to Top Chamber}} \times 100$$

***Note:** During incubation with Cell Dissociation Solution/Cell Dye, gently tap the plate on the side to ensure optimal dissociation of the invasive cells that cling to the outer side of the top chamber.

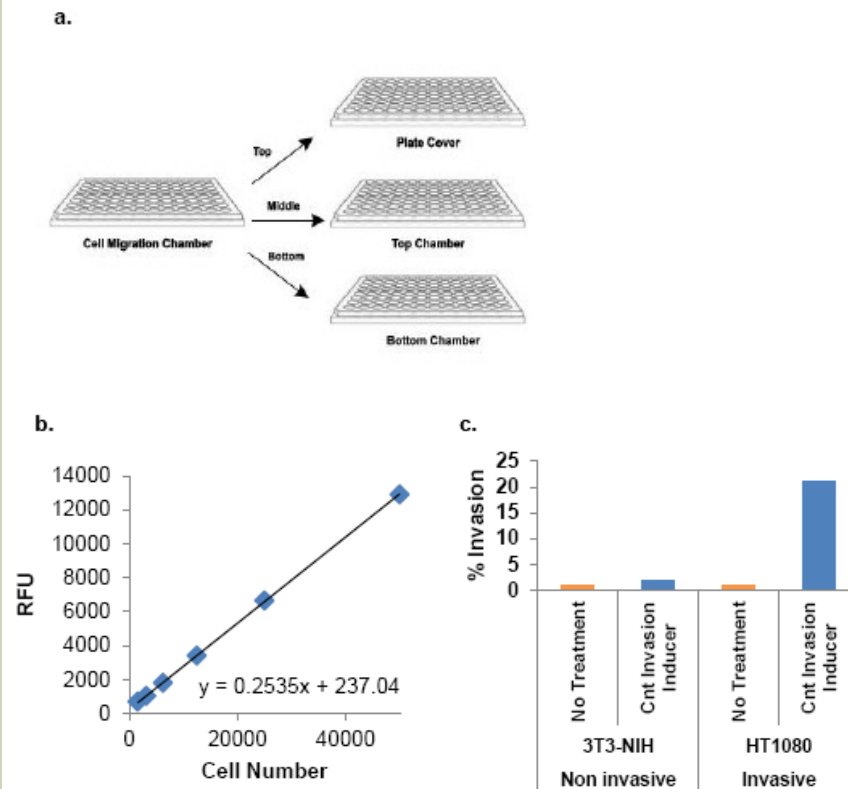


Figure 1: (a) Cell Invasion plate: The cells are added to the Top Chamber and the Control Invasion Inducer or chemoattractant are added to the Bottom Chamber. (b) Standard Curve: HT-1080 cells were harvested, counted and serially diluted to obtain desired cell number. Cells were incubated according to the protocol with Cell Invasion Dye and fluorescence (Ex/Em = 530/590 nm) was measured. (c) Cell Invasion: 3T3-NIH and HT-1080 cells were starved overnight and treated with Control (Cnt) Invasion Inducer or remain untreated (No Treatment). Treatment with Control Invasion Inducer demonstrated a significant increase in invasion of HT 1080 cells as compare to 3T3-NIH control cells.

Intended Use	For in vitro research use only. Not for diagnostic or therapeutic procedures.
Storage & Stability	Store kit at -20°C, protected from light. Briefly centrifuge small vials prior to opening.

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