

3T3-L1 Differentiation Kit

Simple, rapid & convenient method to differentiate 3T3-L1 preadipocytes to adipocytes

Instruction Manual

Catalog Number	PK-CA577-K579												
Description	<p>3T3-L1 cells are derived from mouse 3T3 cells and provide a widely-used fundamental model in the study of adipose physiology and metabolic diseases. They exhibit a fibroblast-like morphology before differentiation but become rounded and accumulate lipid droplets several days after the initiation of differentiation. The accumulated lipid droplets can be visualized by light microscopy. The <i>in vitro</i> differentiated 3T3-L1 adipocytes result in characteristics similar to tissue-derived adipocytes and have been commonly used to study adipogenesis, lipolysis, and metabolic dysfunctions.</p> <p>PromoCell's 3T3-L1 Differentiation Kit provides enough supplements to make 100 ml of differentiation medium and 600 ml of maintenance medium which is sufficient material for 12 x 100 mm culture dishes. The Differentiation Cocktail provides a final concentration of 1.5 µg/ml insulin, 1 µM dexamethasone, 500 µM IBMX, and 1 µM rosiglitazone in the differentiation media.</p>												
Quantity	1 Mix for 100 ml differentiation medium												
Applications	<ul style="list-style-type: none">• Differentiation of 3T3-L1 preadipocytes to adipocytes• Study of obesity, adipogenesis, lipolysis and lipid metabolism												
Sample Type	<ul style="list-style-type: none">• Cell culture: 3T3-L1 cells• Animal tissue: Primary preadipocytes (for primary human preadipocytes see Cat.Nos. C-12730 to C-12733)												
	<table border="1"><thead><tr><th>Components</th><th>Quantity</th><th>Color Code</th></tr></thead><tbody><tr><td>Insulin (1.5 mg/ml)</td><td>0.6 ml</td><td>Green Blue</td></tr><tr><td>Differentiation Cocktail, 1000x (lyophilized)</td><td>1 vial</td><td>Yellow</td></tr><tr><td>DMSO (anhydrous)</td><td>0.5 ml</td><td>Blue</td></tr></tbody></table>	Components	Quantity	Color Code	Insulin (1.5 mg/ml)	0.6 ml	Green Blue	Differentiation Cocktail, 1000x (lyophilized)	1 vial	Yellow	DMSO (anhydrous)	0.5 ml	Blue
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User Supplied Reagents & Equipment	<ul style="list-style-type: none">• Cells grown in 96-well, 6-well, or 100 mm cell culture plate• DMEM, DMEM/F12 (1:1), bovine calf serum, fetal bovine serum (FBS)• Penicillin, streptomycin• 0.22 µM syringe filters• Light Microscope												
Storage and Reagents Preparation	<p>Upon arrival, store kit at -20°C, protected from light. Briefly centrifuge small vials prior to opening. Read the entire protocol before performing the assay.</p> <ul style="list-style-type: none">• Insulin: Ready to use as supplied. Warm to room temperature before use. Aliquot and store at -20°C. Avoid repeated freeze/thaw. Stable for 6 months.• Differentiation Cocktail: Reconstitute in 110 µl DMSO (supplied), making sure the material is completely dissolved. Aliquot and store at -20°C. Avoid repeated freeze/thaw. Stable for 6 months.												
Assay Protocol	<p>1. Cell Culture: Culture 3T3-L1 (ATCC® CL-173TM) in preadipocyte medium consisting of DMEM media with 10% bovine calf serum, 100 units/ml penicillin and 100 µg/ml streptomycin in a humidified incubator at 37°C with 5% CO₂.</p> <p>Notes:</p> <ol style="list-style-type: none">Important: Never allow cultures to become confluent until initiation of differentiation. Change medium every 2-3 days.It is Important to subculture preadipocytes in a medium with 10% bovine calf serum. <p>2. Differentiation Induction: To initiate differentiation, culture cells until confluent. Replace medium with fresh preadipocyte medium and incubate an additional 48 hours. Add 1 µl of Differentiation Cocktail to 1 ml of DMEM/F12 (1:1) with 10% FBS. Make as much differentiation medium as needed. Sterilize with a 0.22 µM syringe filter. Replace preadipocyte medium with differentiation medium. Incubate for 3 days in a humidified incubator at 37°C with 5% CO₂.</p>												

Note:

a. It may be necessary to screen several lots of FBS, as some may be better at differentiation than others.

b. Primary preadipocytes may differentiate better at 10% CO₂.

3. Maintenance: Prepare maintenance medium by adding 1 µl of Insulin to 1 ml of DMEM/F12 (1:1) with 10% FBS. Filter sterilize with 0.22 µM syringe filter. Remove differentiation medium and replace with maintenance medium. Replace medium every 2-3 days. Lipid droplet accumulation will be visible by light microscopy 7-10 days after the addition of differentiation medium.

Note: Enough maintenance medium can be prepared for several medium changes. Store the unused maintenance medium at 4°C.

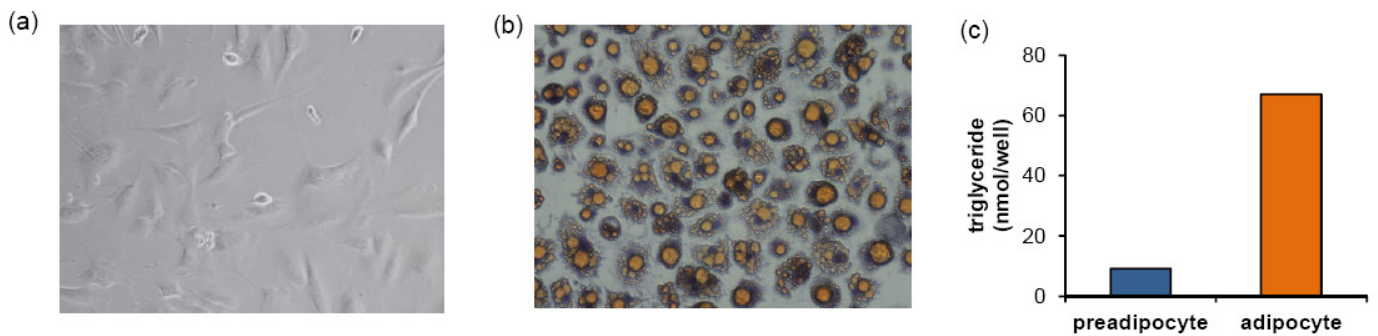


Figure: (a) 3T3-L1 cells before differentiation. (b) 3T3-L1 cells 7 days after the initiation of differentiation and stained with neutral lipid dye Oil Red O (Lipid Staining Kit; Cat.No. PK-CA577-K580). (c) Triglyceride levels in preadipocytes grown in a 96-well cell culture plate and in adipocytes 7 days after differentiation.

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