Product Description

Articular cartilage covers the bones in the major joints of the human body, e.g. knee and hip joints. It is composed of only one specialized cell type, Chondrocytes. Chondrocytes produce and maintain the extracellular matrix of cartilage, i.e. collagen (mostly type II) and proteoglycans. The isolated, differentiated cells express collagen type II; however, during prolonged \textit{in vitro} cultivation, dedifferentiation begins and collagen type II expression declines.

PromoCell Human Chondrocytes (HCH) are produced at PromoCell’s cell culture facility and are available from knee and hip joint cartilage tissue. Shortly after isolation, all PromoCell Human Chondrocytes are cryopreserved at passage 2 (P2) using PromoCell’s proprietary, serum-free freezing medium, Cryo-SFM. Each cryo vial contains more than 500,000 viable cells after thawing. Proliferating cell cultures are made from 500,000 cryopreserved cells that have been thawed and cultured for three days at PromoCell.

Quality Control

Rigid quality control tests are performed for each lot of PromoCell Human Chondrocytes. They are tested for cell morphology, adherence rate, and cell viability. Furthermore, each lot is routinely tested for the capacity to differentiate in 3D spheroids. Growth performance is tested through multiple passages up to 10 population doublings (PD) under culture conditions without antibiotics and antifungics.

In addition, all cells have been tested for the absence of HIV-1, HIV-2, HBV and HCV, and microbial contaminants (fungi, bacteria, and mycoplasma).

A detailed certificate of analysis (CoA) for each lot can be downloaded at: www.promocell.com/coa

Intended Use

PromoCell Human Chondrocytes are for \textit{in vitro} research use only and not for diagnostic or therapeutic procedures.

Warning

Although tested negative for HIV-1, HIV-2, HBV, and HCV, the cells - like all products of human origin - should be handled as potentially infectious. No test procedure can completely guarantee the absence of infectious agents.

Follow appropriate safety precautions!

After delivery, start immediately with the protocol for cryopreserved cells (see page 2) or the protocol for proliferating cells (see page 3).
Start immediately after delivery.
Use aseptic techniques and a laminar flow bench.

Protocol for Cryopreserved Cells

Straight after arrival, store the cryopreserved cells in liquid nitrogen, or seed them immediately.

Note: Storage at - 80°C is not sufficient for cell preservation and causes irreversible cell damage.

1. Prepare the medium
   Calculate the needed culture surface area according to the plating density (see page 5) and the lot-specific cell numbers stated on the certificate of analysis. Fill the appropriate volume of PromoCell Growth Medium (at least 9 ml per vial of cells) in cell culture vessels. Place the vessels in an incubator (37°C, 5% CO₂) for 30 minutes.

2. Thaw the cells
   Remove the cryovial from the liquid nitrogen container and immediately place it on dry ice - even for short transportation. Under a laminar flow bench, briefly twist the cap a quarter turn to relieve pressure, then re-tighten. Immerse the vial into a water bath (37°C) just up to the screw cap for 2 minutes. Ensure that no water enters the thread of the screw cap.

3. Disinfect the vial and seed the cells
   Thoroughly rinse the cryovial with 70% ethanol under a laminar flow bench. Then, aspirate the excess ethanol from the thread area of the screw cap. Open the vial and transfer the cells to a cell culture vessel containing the prewarmed medium from step 1.

4. Incubate the cells
   Place the vessel in an incubator (37°C, 5% CO₂) for cell attachment. Replace the medium after 16 - 24 hours and every two to three days thereafter. The cells should be subcultured, according to the subcultivation protocol (see page 4), once they have reached 70 - 90% confluency.
Start immediately after delivery.
Use aseptic techniques and a laminar flow bench.

Protocol for Proliferating Cells

1. Incubate the cells
   Unpack the culture vessel, do not open the lid, and immediately place it in an incubator (37°C, 5% CO₂) for 3 hours to allow the cells to recover from the transportation.

2. Replace the transport medium
   Carefully open the vessel, rinse the inner side of the lid with 70% ethanol, and let air dry. Aspirate the transport medium from the vessel. Add 10 ml of the appropriate PromoCell Cell Growth Medium.

3. Check and incubate the cells
   Check the cell density. Open the lid half a turn and place the vessel in an incubator (37°C, 5% CO₂). The cells should be subcultured, according to the subcultivation protocol (see page 4), once they have reached > 70% confluency. Change the medium every two to three days.
Use aseptic techniques and a laminar flow bench.

Subcultivation Protocol

1. Prepare the reagents and wash the cells
   Place the PromoCell DetachKit at room temperature for at least 30 minutes to adjust the temperature of the reagents. Carefully aspirate the medium from the culture vessel. Add 100 µl Hepes BSS Solution per cm² of vessel surface to wash the cells and agitate the vessel carefully for 15 seconds.

2. Detach the cells
   Carefully aspirate the Hepes BSS from the culture vessel. Add 100 µl Trypsin/EDTA Solution per cm² of vessel surface. Note: We recommend detaching the cells at room temperature. Close the vessel and examine the cells under a microscope. When the cells start to detach, gently tap the side of the vessel to loosen the remaining cells.

3. Neutralize the trypsin and harvest the cells
   Add 100 µl Trypsin Neutralization Solution per cm² of vessel surface and gently agitate. Carefully aspirate the cell suspension and transfer it to a centrifugation tube. Spin down the cells for 3 minutes at 220 x g.

4. Incubate the cells
   Discard the supernatant (step 1), add 1 ml of the appropriate PromoCell Cell Growth Medium (step 2), and resuspend the cells by carefully pipetting up and down. Plate the cells according to the recommended seeding density in new cell culture vessels containing PromoCell Cell Growth Medium prewarmed to 37°C. Place the vessels in an incubator (37°C, 5% CO₂) and change the media every two to three days.
### Specifications

<table>
<thead>
<tr>
<th>Product</th>
<th>Recommended Culture Media*</th>
<th>Plating Density</th>
<th>Passage after Thawing</th>
<th>Marker</th>
<th>Population Doublings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human Chondrocytes (HCH)</td>
<td>C-27101</td>
<td>10,000 - 20,000 cells per cm²</td>
<td>P2</td>
<td>Differentiation tested</td>
<td>&gt; 10</td>
</tr>
</tbody>
</table>

### Related Products

<table>
<thead>
<tr>
<th>Product</th>
<th>Size</th>
<th>Catalog Number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chondrocyte Growth Medium (Ready-to-use)</td>
<td>500 ml</td>
<td>C-27101</td>
</tr>
<tr>
<td>Chondrocyte Basal Medium</td>
<td>500 ml</td>
<td>C-27111</td>
</tr>
<tr>
<td>Chondrocyte Basal Medium, phenol red-free</td>
<td>500 ml</td>
<td>C-27115</td>
</tr>
<tr>
<td>Chondrocyte Growth Medium SupplementMix</td>
<td>for 500 ml</td>
<td>C-39635</td>
</tr>
<tr>
<td>DetachKit</td>
<td>30 ml, 125 ml, 250 ml</td>
<td>C-41200, C-41210, C-41220</td>
</tr>
<tr>
<td>Cryo-SFM</td>
<td>30 ml, 125 ml</td>
<td>C-29910, C-29912</td>
</tr>
<tr>
<td>HCH Pellet</td>
<td>1 million cells per pellet</td>
<td>C-14070</td>
</tr>
</tbody>
</table>

*The catalog numbers in this table are for media in ready-to-use packaging.*